



RESEARCH ARTICLE

**Quercetin-3-O- α -L-Rhamnopyranosyl-(1-6)- β -D-Glucopyranoside Isolated from
Bougainvillea Glabra**

Sahu N*, Saxena J

*Department of Chemistry, Sarojini Naidu Government Girls (Post Graduate Autonomous) College,
Shivaji Nagar, Bhopal, M.P, India.*

Manuscript No: IJPRS/V3/I1/00063, Received On: 06/02/2014, Accepted On: 15/02/2014

ABSTRACT

Flavonoids were isolated from *Bougainvillea Glabra* using column and thin layer chromatography separation techniques. By comparing their, UV, $^1\text{H-NMR}$, $^{13}\text{C-NMR}$ spectral data proved it to be quercetin – 3 – O – α - L-rhamnopyranosyl - (1 - 6) – β – D – glucopyranoside.

KEYWORDS

Antioxidants Activity, Quercetin – 3 – O – α - L-rhamnopyranosyl - (1 - 6) – β – D – Glucopyranoside, $^{13}\text{C-NMR}$, $^1\text{H-NMR}$

INTRODUCTION

Most flowering ornamental plants are utilized more for their beauty as they radiate different colors to the surroundings. When the flowers bloom, they serve their purpose; however, when they wilt, they just fall off as trash. Before this happens, these flowers could still be of use as pharmaceuticals or nutraceuticals. Several studies show that flowers have a wide array of secondary metabolites of medicinal value, offering antioxidant, antifungal, antibacterial and cytotoxic activities.^{1,2,3}

Secondary metabolites from plants have important biological and pharmacological activities, such as anti-oxidative, anti-allergic, antibiotic, hypoglycemic and anti-carcinogenic.^{4,5,6}

Nutritionists have shown an increased interest in plant antioxidants which could be used in

unmodified form as natural food preservatives to replace synthetic substances.⁷

MATERIALS AND METHOD

Plant Material

The plant materials (flowers) were collected from Guru Govind Singh nursery, Bhopal. The flowers were dried under shade at room temperature. The flowers were powder and stored in sterile container for the further use. The dried powder was then treated with hydro-alcoholic solution (50 ml each) in a soxhlet apparatus for 72hours according to successive solvent extraction. Then the solvent was removed by evaporation and brownish colour semisolid crude extract was obtained

Phytochemical Screening of Plant Extract

A small amount of the dry extract was used for the phytochemical tests⁸ for compounds which include alkaloids, flavonoids, tannins, saponins, glycosides, phenol and terpenoids while steroids, coumarin and cardiac glycosides are absent in all the crude extracts.

***Address for Correspondence:**

Neha Sahu

Department of chemistry,
Sarojini Naidu Government Girls College,
Shivaji Nagar, Bhopal, M.P, India.

E-Mail Id: nehasa88@gmail.com

Isolation and Identification

The isolation of the methanol extract was subjected to column chromatography with silica gel (60-120 mesh) as the stationary phase. The charged column was then eluted with different mobile phases with gradual increase in polarity. The fractions were collected and the solvent recovered by simple distillation. All the concentrated fractions were subjected to TLC for the identification of the desired bands

TLC was performed on the 20 \times 20 cm plates precoated with silica gel (Sigma Aldrich Co., India). TLC analysis of acetic acid extracts were performed using 2 propanol-ethanol-acetic acid (6:7:1) developing solvent systems Based on the Rf-value, number of fractions were obtained and the one with good resolution was visualized under ultraviolet (UV) light, indicating that it was a pure compound, was selected.

Supporting evidence for the structure of the flavone and glycoside is provided by the UV and NMR (125 MHz, DMSO) Spectral data that were recorded on a Bruker AMX 400 NMR spectrometer. Chemical shifts were referenced to the respective residual solvent peaks and the values were recorded in δ .

RESULTS AND DISCUSSION

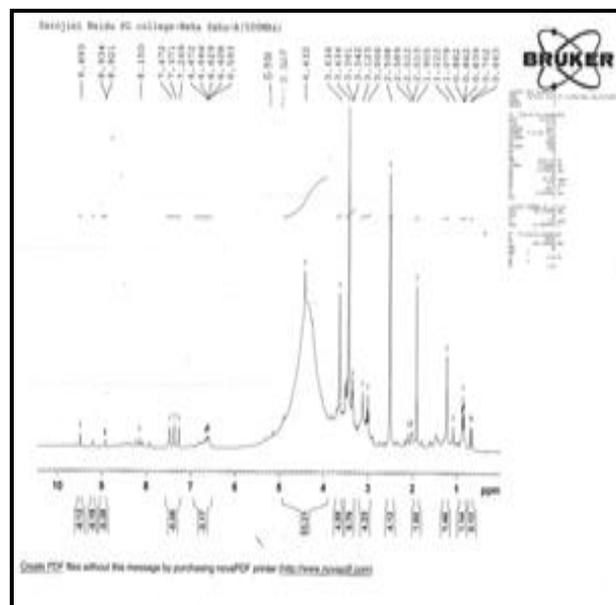
Chemical Constituents

The flowers of *Bougainvillea glabra* have been found to contain Quercetin-3-O- α -L-rhamnopyranosyl-(1-6)- β -Dglucopyranoside.

The UV spectrum of the aglycone exhibited two major peaks at 278 nm (band-I) and 232 nm (band-II), to reveal a flavonoid skeleton. The ^1H and ^{13}C NMR spectra (Table 1) showed the expected signals in the aromatic region for the quercetin moiety in the flavonoids. The ^{13}C NMR data indicated that were 27 carbons in this structure, 15 of which were typical for a flavone skeleton, while others were assigned to glycoside. In the ^1H - NMR spectrum, A-ring protons at C-6 and C-8 appear separately at δ 5.90 ppm and δ 6.593 ppm respectively. The signal at δ 7.37 ppm corresponds to the protons at C-2| and C-6|. The protons at C-5| appear at δ

6.67 ppm. The two signals were observed in the region characteristic for anomeric protons of sugars. Doublets at δ 5.32 ppm were assigned to glucose β -linked to the aglycone. Signals at δ 4.43 ppm corresponded to the anomeric proton of α - linked rhamnose¹⁵. The methyl protons of the sugar rhamnose appear at δ 0.88 ppm and rest of the sugar protons appear in the range δ 3.00 - 3.36 ppm.⁹

In ^{13}C NMR signals corresponding to the anomeric carbon of glucose were found at 73.21 ppm and those corresponding to rhamnose were seen at 84.22 ppm. The attachment of the rhamnose to C-6 of the glycosyl moiety was evidenced by the downfield shift of the glycosyl C-6 carbon resonance to δ 60.11 ppm and accompanying up field shift of the resonances of the adjacent carbons C-5 to 73.08 ppm. The chemical shift values of all the recorded sugar carbon resonances confirmed the pyranose form of the two sugar moieties in the Quercetin-3-O- α -L-rhamnopyranosyl-(1-6)- β -glucopyranoside (Figure 1). The sugar moiety was proved to be acylated at C-3 of the aglycone as deduced from the correlation between the anomeric proton at δ ppm 4.43 and the C-3 at δ ppm84.2. By comparing their UV, ^1H NMR and ^{13}C -NMR data it was proved to be Quercetin-3- O- α -L-rhamnopyranosyl-(1-6)- β -D-glucopyranoside with those reported for similar compound.¹⁰



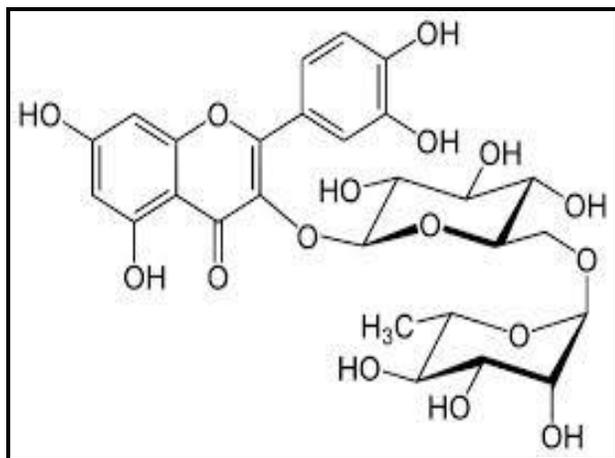


Figure 1: Quercetin-3- O- α -L-rhamnopyranosyl-(1-6)- β -D-glucopyranoside

ACKNOWLEDGEMENT

The author expresses gratitude Prof. Dr. Jyoti Saxena Department of chemistry, S.N.G.G.C. College Bhopal, CMBT Laboratory and SIRT pharmacy department Bhopal for UV-Vis analysis facility and kind support. And IISER, Bhopal for NMR & H-NMR facility.

REFERENCES

- Shindo, K., Saito, E., Sekiya, M., Matsui, T., & Koike, Y. (2008). Antioxidative activity of the flower of *Torenia fournieri*. *Journal of natural medicines*, 62(2), 247-248.
- Nakamura, M., Ishibashi, M., Okuyama, E., Koyano, T., Kowithayakorn, T., Hayashi, M., & Komiyama, K. (2000). Cytotoxic pregnanes from leaves of *Adenium obesum*. *Journal of Natural Medicine*, 54(3), 158-159.
- Umamaheswari, A., Shreevidya, R., & Nuni, A. (2008). *In vitro* antibacterial activity of *Bougainvillea spectabilis* leaves extracts. *Advances in biological Research*, 2(1-2), 1-5.
- Borneo, R., León, A. E., Aguirre, A., Ribotta, P., & Cantero, J. J. (2009). Antioxidant capacity of medicinal plants from the Province of Córdoba (Argentina) and their *in vitro* testing in a model food system. *Food Chemistry*, 112(3), 664-670.
- Katalinic, V., Milos, M., Kulisic, T., & Jukic, M. (2006). Screening of 70 medicinal plant extracts for antioxidant capacity and total phenols. *Food chemistry*, 94(4), 550-557.
- Mulabagal, V., & Tsay, H. S. (2004). Plant cell cultures-an alternative and efficient source for the production of biologically important secondary metabolites. *Int J Appl Sci Eng*, 2(1), 29-48.
- Chien-Chang, S., Yuan-Shiun, C., & Li-Kang, H. (1993). Nuclear Magnetic Resonance Studies of 5,7-Dihydroxyflavonoids. *Phytochem*, 34, 843-845.
- Harborne, J. B. (1993). *The flavonoids advances in research since 1986* (Vol. 4). CRC Press., Williams CA, Harborne JB, Eds., Chapman & Hall, Cambridge, UK, 337 -385.
- Rajamanickam, M., Kalaivanan, P., & Sivagnanam, I. (2013). Antibacterial and Wound Healing Activities of Quercetin-3-OAL-Rhamnopyranosyl-(1-6)- β -D-Glucopyranoside Isolated from *Salvia leucantha*. *International Journal of Pharmaceutical Sciences Review & Research*, 22(1). 77-84.
- Markham, K. R., Ternai, B., Stanley, R., Geiger, H., & Mabry, T. J. (1978). Carbon-13 NMR studies of flavonoids—III: Naturally occurring flavonoid glycosides and their acylated derivatives. *Tetrahedron*, 34(9), 1389-1397.
- Agarwal, A., Gupta, S., & Sharma, R. K. (2005). Role of oxidative stress in female reproduction. *Reprod Biol Endocrinol*, 3(28), 1-21.